In the Specification

Please insert the following paragraphs at page 7, following line 27:

Brief Description of the Sequences

SEQ ID NO:1 is a forward primer used according to the subject invention.

SEQ ID NO:2 is a reverse primer used according to the subject invention.

SEQ ID NO:3 is a forward primer used according to the subject invention.

SEQ ID NO:4 is a reverse primer used according to the subject invention.

SEQ ID NO:5 is a forward primer used according to the subject invention.

SEQ ID NO:6 is a reverse primer used according to the subject invention.

SEQ ID NO:7 is the nucleotide sequence for the primer DNR5'.

SEQ ID NO:8 is the nucleotide sequence for the primer DNR3'.

SEQ ID NO:9 is the nucleotide sequence for the primer DH10Bacinttn7destroybyamp5'.

SEQ ID NO:10 is the nucleotide sequence for the primer DH10Bacinttn7destroybyamp3'.

SEQ ID NO:11 is the nucleotide sequence for the oligonucleotide 5'UNIV.

SEQ ID NO:12 is the nucleotide sequence for the oligonucleotide 3'UNIV.

SEQ ID NO:13 is the nucleotide sequence for the oligonucleotide 5'AVD.

SEQ ID NO:14 is the nucleotide sequence for the oligonucleotide 5'AVD2.

SEQ ID NO:15 is the nucleotide sequence for the oligonucleotide 3'AVD.

SEQ ID NO:16 is the nucleotide sequence for the oligonucleotide 5'EGFP.

SEQ ID NO:17 is the nucleotide sequence for the oligonucleotide 3'EGFP.

Please replace the paragraph at page 14, line 36, through page 15, line 5 with the following:

In order to construct a general baculovirus vector for capsid display, the region corresponding to nucleotides (nt) 469-1506 of vp 39 (Genbank:M22978) was amplified from the purified bacmid DNA (Luckow *et al*, J. Virol. 67, 4566-4579, 1993) by polymerase chain reaction (PCR). The

forward primer was 5' - TT GAA AGA TCT GAA TTC *ATG CAC CAC CAT CAC* CAT CAC GGA TCC GGC GGC GGC GGC TCG **GCG GCT AGT GCC CGT GGG T** - 3' (SEQ ID NO:1; specific sequence for nt 469-486 of vp39 gene in bold; *Bgl*II, *Eco*RI, *Bam*HI, sites underlined; 6 x Histidine tag with start codon in italics); the reverse primer was 5' -TT CTG GGT ACC GCt tta *ATG GTG ATG ATG GTG GTG* TCT AGA GCt tta ACT AGT **GAC GGC TAT TCC TCC ACC** - 3' (SEQ ID NO:2; specific sequence for nt 1489-1506 of vp39 gene in bold; *Kpn*I, *Xba*I and *Spe*I sites underlined; 6 X Histidine tag in italics; stop codon in small caps). PCR was performed essentially as described by Airenne *et al*, Gene 144:75-80, 1994, except annealing was set to 58°C. Amplified fragment was digested with *Bgl*II and *Kpn*I enzymes and purified as described in Airenne *et al*, *supra*. The purified PCR product was cloned into *Bam*HI+*Kpn*I-digested pFastBAC1 vector (Invitrogen, Carlsbad, USA). The resulted plasmid was named as pBACcap-1. The nucleotide sequence was confirmed by sequencing (ALF; Amersham Pharmacia Biotech, Uppsala, Sweden).

Please replace the paragraph at page 15, lines 7-18, with the following:

cDNA encoding EGFP (enhanced green fluorescent protein) was amplified from the pEGFP-N1 plasmid (Genbank:U55762, Clontech, Palo Alto, USA) by PCR and cloned into the pBACcap-1. Two sets of primers were used to enable EGFP fusion both to N- and C-terminal ends of the vp39. For the N-terminal fusion, the forward primer was 5' - CGG GAT *GAA TTC* **GTC GCC ACC ATG GTG AGC AAG GGC GAG GAG** - 3' (SEQ ID NO:3; specific sequence for nt 670-699 of pEGFP-N1 in bold; *Eco*RI site in italics), and the reverse primer 5' - GCG GCC *GGA TCC* **CTT GTA CAG CTC GTC CAT GCC** - 3' (SEQ ID NO:4; specific sequence for nt 1375-1395 of pEGFP-N1 in bold; *Bam*HI site in italics). The amplified fragment which corresponded to nt 670-1395 of pEGFP-N1 was cloned into *Eco*RI/*Bam*HI site of the *Spe*I/*Xba*I-deleted pBACcap-1. The resulting plasmid was named pEGFPvp39.

Please replace the paragraph at page 15, lines 19-27, with the following:

For the C-terminal version, the forward primer was 5' - GTC GCC *ACT AGT* **GTG AGC AAG GGC GAG GAG CTG** -3' (SEQ ID NO:5; specific sequence for nt 682-702 of pEGFP-N1 in bold; *Spe*I site in italics), and the reverse primer 5' - AGA GTC *ACT AGT* GCt **tta CTT GTA CAG CTC GTC CAT GCC** - 3' (SEQ ID NO:6; specific sequence for nt 1375-1398 of pEGFP-N1 in bold; *Spe*I site in italics; stop codon in small caps). The amplified fragment which corresponded to nt 682-1398 of pEGFP-N1 was cloned into *Spe*I site of the pBACcap-1. The resulting plasmid was named pvp39EGFP. The nucleotide sequences were confirmed by sequencing (ALF).

Please replace the paragraph at page 20, line 26 through page 21, line 11, with the following:

The modified donor vector was constructed by replacing the Ampicillin resistance gene in pFastbac1 vector with *Bacillus subtilis* levansucrase gene (*SacB*) from pDNR-LIB vector. In practice, pFastbac1 vector was cut by *Bsp*HI restriction enzyme, and the linear vector backbone was purified by gel electrophoresis. The *SacB* expression cassette was obtained from pDNR-LIB by polymerase chain reaction (PCR) with the primers DNR5': 5' B GTTATTCATGAGATCT**GTCAATGCCAATAGGATATC** B 3' (SEQ ID NO:7; sequence for nt 1263-1282 of pDNR-LIB in bold; *Bsp*HI and *Bgl*II sites underlined), DNR3': 5' B TTAGGTCATGA**ACATATACCTGCCGTTCACT** B 3' (SEQ ID NO:8; sequence for nt 3149-3179 of pDNR-LIB in bold; *Bsp*HI site underlined). PCR was performed essentially as described by Airenne *et al* (1994), *supra*, except that annealing was carried out at 58□C and EXT DNA polymerase (Finnzymes, Helsinki, Finland) was used for amplification. The amplified fragment was digested with *Bsp*HI and purified as described in Airenne *et al*, (1994), *supra*. The purified PCR product was cloned into a *Bsp*HI-digested pFastbac1 vector (Invitrogen, Carlsbad, USA) for orientation shown in Figure 2. The resulting plasmid was named pBVboost. The *SacB#3* cassette nucleotide sequence was confirmed by DNA sequencing (ALF; Amersham Pharmacia Biotech, Uppsala, Sweden).

Please replace the paragraph at page 21, lines 13-26, with the following:

In order to block the cryptic *att*Tn7 site in DH10Bac, pBVboost was cut by *Bse*RI/*Avr*II. The excised gentamycin resistance was substituted by ampicillin resistance cassette (ARC) from pFastbac1. The *ARC* was obtained by PCR with the primers DH10Bacinttn7destroybyamp5': 5'-AAATATGAGGAGTTACAATTGCTAATTAATTAAT**TCGGGGAAATGTGCGCGGAA** B 3' (SEQ ID NO:9; sequence for nt 471-490 of pFastbac1 in bold; *Bse*RI site underlined), DH10Bacinttn7destroybyamp3': 5' B CTTGGTCCTAGGATTA**CCAATGCTTAATCAGTG** B 3' (SEQ ID NO:10; sequence for nt 1430-1449 of pFastbac1 in bold; *Avr*II site underlined). The PCR was performed as described above. The amplified fragment was digested with *Bse*RI/*Avr*II and purified as above. The purified PCR product was cloned into a *Bse*RI/*Avr*II-digested pBVboost. The resulting plasmid was named pBVboostΔamp. The nucleotide sequence of Ampicillin cassette was confirmed by DNA sequencing (ALF; Amersham Pharmacia Biotech, Uppsala, Sweden).